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Extracellular Superoxide Dismutase and Risk of COPD

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ABSTRACT

Chronic obstructive pulmonary disease (COPD) is a major public health problem worldwide. COPD is strongly related to cigarette smoke exposure, but not all smokers develop the disease. It is thought that COPD progresses slowly over time stimulated by environmental exposures, including free radicals from cigarette smoke, which ultimately establish chronic inflammation and result in a progressive destruction of lung tissues. COPD is known to occur in family clusters, which has prompted interest in determining genetic risk factors for the disease. Several genetic studies have identified an association between extracellular superoxide dismutase (ECSOD) polymorphisms and risk for developing COPD. ECSOD is an antioxidant protein that scavenges superoxide free radicals from cigarette smoke and protects the lungs from free radical damage and chronic inflammation.

INTRODUCTION

Chronic Obstructive Pulmonary Disease (COPD) is the fourth leading cause of death in the world and is a major public health problem due to its associated disability and consumption of health care resources. It is primarily related to tobacco smoke exposure, although other inhalational exposures appear to increase the risk of disease. However, not all smokers will develop COPD. A recent study found that only 25% of smokers developed COPD in a Danish population (1). This suggests that individuals have varying susceptibility to develop the disease.

COPD prevalence rises with increasing age and is known to occur in family clusters. Increased risk of COPD in smokers' relatives and recognition of a specific genetic mutation, alpha 1 antitrypsin deficiency, that leads to COPD, have excited interest in defining the genetic risk factors for the disease.

Keywords: COPD, ECSOD, Genetic Association, free radicals *Both co-authors contributed equally. Correspondence to Rebecca Oberley-Deegan Department of Medicine National Jewish Health Denver, Colorado 80206, USA Phone: 303-398-1547 Fax: 202-270-2249 e-mail: oberleyr@jhealth.org Several genetic studies have separately identified an association between extracellular superoxide dismutase (ECSOD or SOD3) polymorphisms and altered risk of COPD (2, 3). ECSOD is an antioxidant and anti-inflammatory protein found in high concentrations in both lung tissue and in the lung lining fluids (4). It has been hypothesized by many that COPD progression involves both the release of free radicals and redox sensitive proteases that result in small airway inflammation, fibrosis and alveolar wall destruction.

Pathophysiology of COPD

COPD is a clinical syndrome defined by chronic expiratory airflow obstruction leading to exercise intolerance and dyspnea. However, a comprehensive description of the disease remains a work in progress. Classic descriptions of chronic bronchitis and emphysema define polar aspects of COPD disease phenotypes; however, the majority of COPD patients display mixed patterns of airway disease and emphysematous loss of lung parenchyma. High resolution chest CT scans of COPD subjects demonstrate various combinations of large and small airway disease leading to air trapping and variable patterns and degrees of emphysema.

The time course and progression of COPD includes: eigarette smoke or environmentally induced airway inflammatory disease that leads to chronic cough and sputum production. There is a significant gap of years between smoking exposure and the development of clinical disease. Ultimately, immune defenses are

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activated in response to chronic inflammatory cytokines and recurrent infections. Recurrent infections in COPD patients are associated with alterations in immune mechanisms and a role for both macrophages and neutrophils has been defined in COPD pathogenesis (5). The unique characteristic of patients with COPD is that they develop persistent chronic inflammation that is not successfully controlled. These inflammatory events break down lung collagen by chronic activation of local proteases. The results of these chronic inflammatory events are changes in alveolar size and lung compliance producing emphysema (6), and chronic airway inflammation.

Cigarette smoking, COPD and free radicals in the lung

The lung has an extensive surface area that is exposed to environmental irritants, such as cigarette smoke, which causes free radical production. Each puff of cigarette smoke has been reported to contain up to 10¹⁴ free radical molecules as well as 4700 chemicals (7). Some of these chemicals are short term free radicals such as superoxide (O_2^{-}) and nitric oxide (NO) and others are long acting free radicals such as semiquinones (8). Cigarette smoke enhances recruitment of inflammatory cells to the lung (9). Inflammatory cells, such as activated neutrophils and macrophages, can produce large amounts of reactive oxygen species (ROS), mainly through the NADPH oxidase system (10). It has been hypothesized that the ROS released by inflammatory cells recruited to sites of injury, cause extensive tissue damage which leads to chronic inflammation (11). ROS release by neutrophils and macrophages not only damages surrounding tissues, it can also directly damage/inactivate antioxidant enzymes (11). ROS from activated neutrophils can cause proteolysis of the antioxidant, ECSOD, rendering it inactive (11). Thus, chronic free radical production can inhibit the activity of the very enzymes released to protect the body from free radical damage.

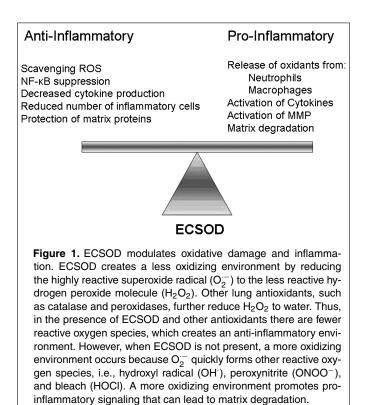
Free radicals and reactive species that generate oxidative stress are short lived; thus, oxidative stress is identified by measuring end products of free radical reactions that have already occurred. These oxidative stress markers include lipid peroxidation (isoprotanes), protein carbonyls, nitrotyrosine formation, glutathione levels and DNA damage. COPD has been associated with increased isoprostanes and lipid peroxidation (12–15), and elevated nitric oxide production and nitrotyrosine formation (16, 17), supporting a role for oxidative stress in the disease. Moreover, these markers are elevated even in mild COPD (18, 19). Leukocytes from COPD subjects have increased generation of superoxide, increased SOD activity, increased protein carbonyls and increased glutathione levels as compared to controls (20).

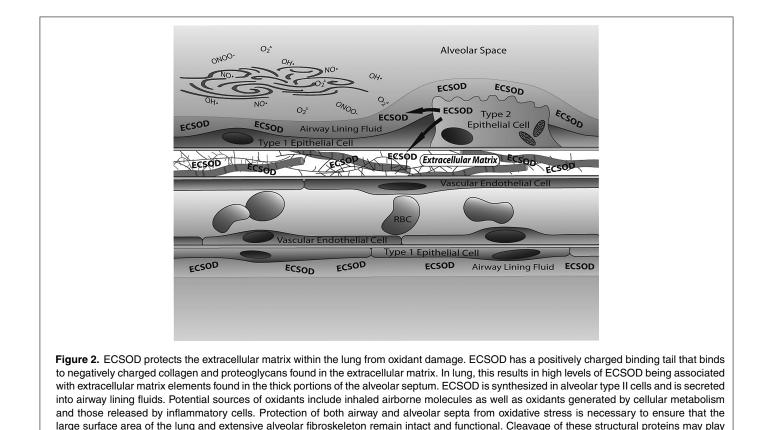
Lung inflammation persists even after a person stops smoking (21). One study found that for an ex-smoker, it takes up to 3 years of not smoking for lung macrophage numbers to decrease to the levels of a never smoker (9). A recent prospective study found that oxidative stress persists in the lungs for months after smoking cessation (22). Another study demonstrated that respiratory bronchiolitis, a common inflammatory lesion of the respiratory bronchioles associated with smoking, can occur or persist well after a person has stopped smoking. Respiratory bronchiolitis was found in 42% of patients who had quit smoking for 3 years and 33% who had quit for 5 years (21). Hogg et al. found increased lymphocyte numbers in airways of patients with severe COPD that had not smoked for an average of 9 years (23). Taken together these data indicate that smoking causes chronic inflammation and oxidative stress in the lung and that both persist well after a person has stopped smoking.

Cigarette smoke can overwhelm the capacity of lung antioxidant defenses and lead to chronic oxidative stress and inflammation in the respiratory system (24, 25). Oxidative stress in the lung may also be perpetuated by recurrent infections with excess accumulation of inflammatory cells.

ECSOD background

The lung has several lines of defense to protect against oxidants, pollutants, and irritants, such as cigarette smoke. One important line of defense is the production of antioxidant enzymes (4). Superoxide dismutases are powerful antioxidant enzymes that reduce the superoxide radical to a less reactive hydrogen peroxide molecule (26). There are two superoxide dismutases localized in lung cells (MnSOD and CuZnSOD) and one superoxide dismutase primarily localized in the extracellular space of the lung (ECSOD, Figure 1) (26–28). Other lung antioxidant enzymes such as catalase, glutathione peroxidases, and the thioredoxin/peroxiredoxin and glutaredoxin families of enzymes further contribute to the scavenging of hydrogen peroxide to water (4). Together these antioxidant enzymes work to protect the lung from oxidatative stress.





The major SOD in extracellular fluids is ECSOD (27). EC-SOD is found in high abundance in the lung, the fluid lining the lung and in the vasculature (29, 30). In particular, ECSOD is located in the lung extracellular matrix, at airway epithelial cell junctions, in the lining of vessels and at the surface of airway smooth muscles (29, 30). ECSOD is a 135,000 mw tetrameric glycoprotein that has a N-terminal signal peptide for secretion out of the cell, a copper/zinc containing activity domain and a C-terminal heparin binding tail (31). The positively charged heparin binding tail allows ECSOD to bind to negatively charged extracellular matrix elements and to endothelial cells (32). The distribution of ECSOD in extracellular compartments indicates that the enzyme plays a critical role in protecting extracellular matrix proteins from free radical damage and potentially protecting this tissue from the progression of chronic inflammation (Figure 2).

a role in the development of emphysema.

ECSOD function in lung

ECSOD is important in protecting the lung from free radical damage and in controlling inflammation (33). ECSOD ameliorates a wide range of lung injuries. ECSOD protects mice from asbestos-induced lung injury (34). Asbestos exposed mice lacking ECSOD have a greater inflammatory response, more fibrosis, and more oxidative damage as compared to exposed wild-type mice (35). In another lung injury model, hyperoxia, toxicity is reduced by high levels of ECSOD (36). In that particular model, animals that overexpress ECSOD had significantly

lower mortality rates as compared to wild type animals and had fewer inflammatory cells in their bronchial alveolar lavage fluid as compared to wild-type mice (36). Recently, Gongora, et al. found that conditional ECSOD knock-out in mature animals leads to acute lung injury at ambient levels of oxygen (37).

ECSOD has also been shown to protect against bleomycininduced pulmonary fibrosis. Mice that overexpressed ECSOD had less fibrosis and reduced total lung collagen (38). Finally, ECSOD inhibits inflammation associated with lipopolysaccharide (LPS) exposure (39). ECSOD reduced the number of neutrophils in the lung airways and reduced expression of the inflammatory cytokines, TNF- α and MIP-2 (39).

ECSOD has been shown to play an important role in protecting lung extracellular matrix from inflammation *in vitro* (33). ECSOD binds to a variety of human extracellular matrix proteins via its C-terminal binding domain (4). ECSOD binds to collagen, hyaluronan, and heparan sulfate, all proteins found in high abundance in the lung extracellular matrix (40–42). Under oxidative conditions, the extracellular matrix becomes damaged by free radical production and the proteins that make up the extracellular matrix fragment. Accumulation of these collagen, hyaluronan and heparan sulfate fragments elicits inflammatory responses in the extracellular matrix (40–42). When present, ECSOD binds to extracellular matrix proteins and protects these proteins from fragmentation (40–42). Thus, ECSOD protects the human extracellular compartment from damage due to oxidative stress.

Table 1. Summary of genetic association studies of COPD with ECSOD			
Reference	Study Population	Subjects	Findings
Juul et al. (2006)	Danish Population	n = 9258	ECSOD R213G mutation protects smokers from COPD
Young et al. (2006)	European Decent Population	n = 440	ECSOD R213G mutation protects smokers from COPD
Wilk et al. (2007)	Framingham Heart Study	n = 1578	SNPs near ECSOD gene exhibited association to lung function
Dahl et al. (2008)	Copenhagen City Heart Study	n = 9093	An ECSOD polymorphism (E1/I1) was associated with COPD morbidity
Dahl et al. (2008)	Copenhagen General Population Study	n = 35635	No association of ECSOD (E1/I1) with COPD morbidity was found

ECSOD has been shown to inhibit fibrosis. Epithelial cells and macrophages release transforming growth factor- β (TGF- β), which stimulates fibroblast proliferation and leads to fibrotic lesions (43). ECSOD inhibits the TGF- β signaling pathway and thus prevents fibrosis *in vivo* (44). COPD is a disease characterized by sites of inflammation and fibrosis and ECSOD is critical in the control of both of these processes.

Genetic studies involving ECSOD in lung function and COPD disease

Significant familial aggregation of both spirometric lung function and COPD has been shown in a number of studies (45–48). Several genetic studies have shown that ECSOD polymorphisms are associated with normal and altered lung function. In mice, SOD3 variants were associated with reduced lung function (49). In humans, associations between ECSOD polymorphisms and reduced lung function in children and adults have been observed (2, 50). Genome wide association in the Framingham Heart study showed association of a SNP close to the ECSOD gene with percent predicted FEV1 and percent predicted FVC (51). Arcaroli et al. have shown that certain ECSOD haplotypes reduce lung inflammation and decrease the severity of acute lung injury and mortality (52). Therefore, both animal and human genetic studies have shown that polymorphisms in ECSOD are important in lung function, with specific polymorphisms associated with either an increase or a decrease in lung function.

Five genetic studies have evaluated the relationships of EC-SOD and risk of COPD (Table 1, (53)). In two independent studies, using two different populations, ECSOD polymorphisms have been correlated with a reduced risk of developing COPD (3, 54). In particular, the ECSOD polymorphism, R213G, has been shown to reduce the risk of smokers to develop COPD (3, 54). This polymorphism causes a substitution of an arginine for a glycine at amino acid 213 in the heparin binding tail of ECSOD (Figure 3). The normal heparin binding tail of ECSOD has a cluster of 6 positively charged amino acids, which allows ECSOD to bind to the negatively charged extracellular matrix (Figure 3, (3, 54)). The R213G mutation reduces the positive charge on the binding tail, and markedly alters the protein's affinity for binding to tissues or extracellular matrix. A result of this is that high circulating levels occur for the R231G EC-SOD protein- and it is presumed that this is associated with high amounts in the airway lining fluids where it would be in an ideal position to protect the lung from antioxidant injury induced by

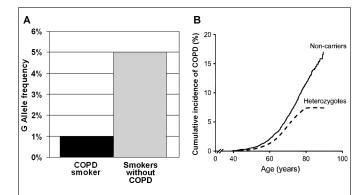
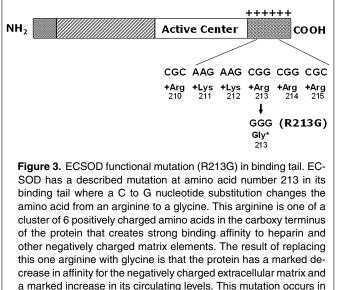


Figure 4. Effect of ECSOD binding tail mutation on COPD risk in smokers. A. This Figure is adapted from Young et al. (54) and shows smokers who have not developed COPD have a significantly higher incidence of the G allele at the R213G locus as compared to smokers who developed COPD. This protection from the effects of smoking is presumed to be related to increased levels of ECSOD secreted into the alveolar lining fluid, and thereby, more effective antioxidant protection from airborne oxidants found in cigarette smoke. B. This figure is adapted from Juul et al. (3). This prospective study followed individuals for an average of 24 years and found that individuals heterozygous for the ECSOD R213G mutation had significantly lower COPD morbidity and morality rates as compared to individuals that were noncarriers for this ECSOD mutation.



4-6% of northern European populations.

inhaled oxidants, such as cigarette smoking. Thus, individuals who smoke and are carriers of this ECSOD polymorphism are at a reduced risk of developing COPD (Figure 4A&B).

SUMMARY

Most people who develop COPD are smokers or former smokers. Smoking increases lung oxidative stress, increases inflammatory cytokines, and causes recruitment of inflammatory cells to the lung. When not controlled, this chronic inflammatory process leads to chronic airway disease, destruction of lung tissue and COPD. COPD is found in family clusters, which suggests that genetics plays an important role in the progression of COPD. Several genetic studies have indicated that the R213G mutation in ECSOD is associated with protection from the development of COPD. The R213G mutation results in high circulating levels of ECSOD and likely high levels in alveolar and airway linking fluids where it could protect the lung interface from the effects of inhaled free radicals in tobacco smoke. ECSOD both scavenges free radicals and controls inflammation associated with COPD.

Both animal models and human genetic studies have demonstrated associations of ECSOD with lung function and development of smoking related diseases. A common ECSOD polymorphism has been shown to reduce the risk of developing COPD. While this polymorphism could account for only a small portion of the smokers who are resistant to developing COPD, it demonstrates the potential importance of ECSOD in protecting the lung from oxidative stress. A relative deficiency or inadequate up-regulation of ECSOD in response to stress may be an important component in enhancing risk for development of COPD.

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Declaration of interest:

The authors report no conflict of interest. The authors alone are responsible for the content and writing of the paper.

REFERENCES

1. Lokke A, Lange P, Scharling H, Fabricius P, Vestbo J. Developing COPD: a 25 year follow up study of the general population. Thorax 2006; 61:935–939.

- Dahl M, Bowler RP, Juul K, Crapo JD, Levy S, Nordestgaard BG. Superoxide dismutase 3 polymorphism associated with reduced lung function in two large populations. Amer J Respir Crit Care Med 2008; 178:906–912.
- Juul K, Tybjaerg-Hansen A, Marklund S, Lange P, Nordestgaard BG. Genetically increased antioxidative protection and decreased chronic obstructive pulmonary disease. Amer J Respir Crit Care Med 2006; 173:858–864.
- Kinnula VL, Crapo JD. Superoxide dismutases in the lung and human lung diseases. Amer J Respir Crit Care Med 2003; 167:1600– 1619.
- 5. Tetley TD. Inflammatory cells and chronic obstructive pulmonary disease. Curr Drug Targets 2005; 4:607–618.
- Yoshida T, Tuder RM. Pathobiology of cigarette smoke-induced chronic obstructive pulmonary disease. Physiol Rev 2007; 87:1047–1082.
- Church DF, Pryor WA. Free-radical chemistry of cigarette smoke and its toxicological implications. Environ Health Perspect 1985; 64:111–126.
- Kirkham P, Rahman I. Oxidative stress in asthma and COPD: antioxidants as a therapeutic strategy. Pharmacol Therapeut 2006; 111:476–494.
- Agius RM, Rutman A, Knight RK, Cole PJ. Human pulmonary alveolar macrophages with smokers' inclusions: their relation to the cessation of cigarette smoking. Br J Exper Pathol 1986; 67:407–413.
- Iles KE, Forman HJ. Macrophage signaling and respiratory burst. Immunol Res 2002;26:95–105.
- McCord JM, Gao B, Leff J, Flores SC. Neutrophil-generated free radicals: possible mechanisms of injury in adult respiratory distress syndrome. Environ Health Perspect 1994;102 Suppl 10:57–60.
- 12. Biernacki WA, Kharitonov SA, Barnes PJ. Increased leukotriene B4 and 8-isoprostane in exhaled breath condensate of patients with exacerbations of COPD. Thorax 2003;58:294–298.
- Paredi P, Kharitonov SA, Leak D, Ward S, Cramer D, Barnes PJ. Exhaled ethane, a marker of lipid peroxidation, is elevated in chronic obstructive pulmonary disease. Amer J Respirat Crit Care Med 2000;162:369–373.
- Pratico D, Basili S, Vieri M, Cordova C, Violi F, Fitzgerald GA. Chronic obstructive pulmonary disease is associated with an increase in urinary levels of isoprostane F2alpha-III, an index of oxidant stress. Amer J Respir Crit Care Med 1998; 158:1709– 1714.
- Rahman I, van Schadewijk AA, Crowther AJ, Hiemstra PS, Stolk J, MacNee W, De Boer WI. 4-Hydroxy-2-nonenal, a specific lipid peroxidation product, is elevated in lungs of patients with chronic obstructive pulmonary disease. Amer J Respirat Crit Care Med 2002;166:490–495.
- Corradi M, Pesci A, Casana R, Alinovi R, Goldoni M, Vettori MV, Cuomo A. Nitrate in exhaled breath condensate of patients with different airway diseases. Nitric Oxide 2003; 8:26–30.
- Ichinose M, Sugiura H, Yamagata S, Koarai A, Shirato K. Increase in reactive nitrogen species production in chronic obstructive pulmonary disease airways. Amer J Respir Critical Care Med 2000; 162:701–706.
- Kinnula VL, Ilumets H, Myllarniemi M, Sovijarvi A, Rytila P. 8-Isoprostane as a marker of oxidative stress in nonsymptomatic cigarette smokers and COPD. Eur Respir J 2007; 29:51–55.
- Rytila P, Rehn T, Ilumets H, Rouhos A, Sovijarvi A, Myllarniemi M, Kinnula VL. Increased oxidative stress in asymptomatic current chronic smokers and GOLD stage 0 COPD. Resp Res 2006; 7:69.
- 20. Nadeem A, Raj HG, Chhabra SK. Increased oxidative stress and altered levels of antioxidants in chronic obstructive pulmonary disease. Inflammation 2005; 29:23–32.
- Fraig M, Shreesha U, Savici D, Katzenstein AL. Respiratory bronchiolitis: a clinicopathologic study in current smokers,

ex-smokers, and never-smokers. Amer J Surg Pathol 2002; 26:647–653.

- Louhelainen N, Rytila P, Haahtela T, Kinnula VL, Djukanovic R. Persistence of oxidant and protease burden in the airways after smoking cessation. BMC Pulmon Med 2009; 9:25.
- Hogg JC, Chu F, Utokaparch S, Woods R, Elliott WM, Buzatu L, Cherniack RM, Rogers RM, Sciurba FC, Coxson HO, Pare PD. The nature of small-airway obstruction in chronic obstructive pulmonary disease. New Engl J Med 2004; 350:2645–2653.
- 24. Bowler RP, Barnes PJ, Crapo JD. The role of oxidative stress in chronic obstructive pulmonary disease. COPD 2004; 1:255–277.
- **25.** Rahman I. The role of oxidative stress in the pathogenesis of COPD: implications for therapy. Treat Respir Med 2005; 4:175–200.
- McCord JM, Fridovich I. Superoxide dismutase. An enzymic function for erythrocuprein (hemocuprein). J Biol Chem 1969; 244:6049–6055.
- 27. Marklund SL. Human copper-containing superoxide dismutase of high molecular weight. *Proceedings of the National Academy of Sciences of the United States of America* 79: 7634–7638, 1982.
- **28.** Weisiger RA, Fridovich I. Mitochondrial superoxide dimutase. Site of synthesis and intramitochondrial localization. J Biol Chem 1973; 248:4793–4796.
- Oury TD, Chang LY, Marklund SL, Day BJ, Crapo JD. Immunocytochemical localization of extracellular superoxide dismutase in human lung. Lab Invest J Tech Meth Pathol 1994; 70:889–898.
- Oury TD, Day BJ, Crapo JD. Extracellular superoxide dismutase in vessels and airways of humans and baboons. Free Rad Biol Med 1996; 20:957–965.
- **31.** Folz RJ, Crapo JD. Extracellular superoxide dismutase (SOD3): tissue-specific expression, genomic characterization, and computer-assisted sequence analysis of the human EC SOD gene. Genomics 1994; 22:162–171.
- 32. Inoue M, Watanabe N, Matsuno K, Sasaki J, Tanaka Y, Hatanaka H, Amachi T. Expression of a hybrid Cu/Zn-type superoxide dismutase which has high affinity for heparin-like proteoglycans on vascular endothelial cells. J Biol Chem 1991; 266:16409– 16414.
- Gao F, Kinnula VL, Myllarniemi M, Oury TD. Extracellular superoxide dismutase in pulmonary fibrosis. Antioxidants Redox Signal 2008; 10:343–354.
- Tan RJ, Fattman CL, Watkins SC, Oury TD. Redistribution of pulmonary EC-SOD after exposure to asbestos. J Appl Physiol 2004; 97:2006–2013.
- Fattman CL, Tan RJ, Tobolewski JM, Oury TD. Increased sensitivity to asbestos-induced lung injury in mice lacking extracellular superoxide dismutase. Free Rad Biol Med 2006; 40:601–607.
- Folz RJ, Abushamaa AM, Suliman HB. Extracellular superoxide dismutase in the airways of transgenic mice reduces inflammation and attenuates lung toxicity following hyperoxia. J Clin Invest 1999; 103:1055–1066.
- 37. Gongora MC, Lob HE, Landmesser U, Guzik TJ, Martin WD, Ozumi K, Wall SM, Wilson DS, Murthy N, Gravanis M, Fukai T, Harrison DG. Loss of extracellular superoxide dismutase leads to acute lung damage in the presence of ambient air: a potential mechanism underlying adult respiratory distress syndrome. *Amer J Pathol* 2008; 173:915–926.
- Bowler RP, Nicks M, Warnick K, Crapo JD. Role of extracellular superoxide dismutase in bleomycin-induced pulmonary fibrosis. *Amer J Physiol* 2002; 282:L719–726.
- 39. Bowler RP, Nicks M, Tran K, Tanner G, Chang LY, Young SK, Worthen GS. Extracellular superoxide dismutase attenu-

ates lipopolysaccharide-induced neutrophilic inflammation. Amer J Respir Cell Mol Biol 2004; 31:432–439.

- Gao F, Koenitzer JR, Tobolewski JM, Jiang D, Liang J, Noble PW, Oury TD. Extracellular superoxide dismutase inhibits inflammation by preventing oxidative fragmentation of hyaluronan. J Biol Chem 2008; 283:6058–6066.
- 41. Kliment CR, Tobolewski JM, Manni ML, Tan RJ, Enghild J, Oury TD. Extracellular superoxide dismutase protects against matrix degradation of heparan sulfate in the lung. Antioxidants Redox Signal 2008; 10:261–268.
- Petersen SV, Oury TD, Ostergaard L, Valnickova Z, Wegrzyn J, Thogersen IB, Jacobsen C, Bowler RP, Fattman CL, Crapo JD, Enghild JJ. Extracellular superoxide dismutase (EC-SOD) binds to type I collagen and protects against oxidative fragmentation. J Biol Chem 2004; 279:13705–13710.
- **43. Cutroneo KR.** TGF-beta-induced fibrosis and SMAD signaling: oligo decoys as natural therapeutics for inhibition of tissue fibrosis and scarring. *Wound Repair Regen* 15 Suppl 1: S54–60, 2007.
- 44. Rabbani ZN, Anscher MS, Folz RJ, Archer E, Huang H, Chen L, Golson ML, Samulski TS, Dewhirst MW, Vujaskovic Z. Overexpression of extracellular superoxide dismutase reduces acute radiation induced lung toxicity. *BMC Cancer* 2005; 5:59.
- **45.** Cotch MF, Beaty TH, Cohen BH. Path analysis of familial resemblance of pulmonary function and cigarette smoking. Amer Rev Respir Dis 1990; 142:1337–1343.
- Cotch MF, Beaty TH, Munoz A, Cohen BH. Estimating familial aggregation while adjusting for covariates. Application to pulmonary function data from black and white sibships. Ann Epidemiol 1992; 2:317–324.
- **47. Rybicki BA, Beaty TH, Cohen BH.** Major genetic mechanisms in pulmonary function. J Clin Epidemiol 1990; 43:667–675.
- Silverman EK, Chapman HA, Drazen JM, Weiss ST, Rosner B, Campbell EJ, O'Donnell WJ, Reilly JJ, Ginns L, Mentzer S, Wain J, Speizer FE. Genetic epidemiology of severe, early-onset chronic obstructive pulmonary disease. Risk to relatives for airflow obstruction and chronic bronchitis. Amer J Respir Crit Care Med 1998; 157:1770–1778.
- Reinhard C, Meyer B, Fuchs H, Stoeger T, Eder G, Ruschendorf F, Heyder J, Nurnberg P, de Angelis MH, Schulz H. Genomewide linkage analysis identifies novel genetic Loci for lung function in mice. Amer J Respir Crit Care Med 2005; 171:880–888.
- Ganguly K, Stoeger T, Wesselkamper SC, Reinhard C, Sartor MA, Medvedovic M, Tomlinson CR, Bolle I, Mason JM, Leikauf GD, Schulz H. Candidate genes controlling pulmonary function in mice: transcript profiling and predicted protein structure. *Physiol Genom* 2007; 31:410–421.
- Wilk JB, Walter RE, Laramie JM, Gottlieb DJ, O'Connor GT. Framingham Heart Study genome-wide association: results for pulmonary function measures. BMC Med Genetics 2007; 8 Suppl 1: S8.
- 52. Arcaroli JJ, Hokanson JE, Abraham E, Geraci M, Murphy JR, Bowler RP, Dinarello CA, Silveira L, Sankoff J, Heyland D, Wischmeyer P, Crapo JD. Extracellular superoxide dismutase haplotypes are associated with acute lung injury and mortality. Amer J Respir Crit Care Med 2009; 179:105–112.
- **53.** Bentley AR, Emrani P, Cassano PA. Genetic variation and gene expression in antioxidant related enzymes and risk of COPD: a systematic review. Thorax 2008; 63:956–961.
- Young RP, Hopkins R, Black PN, Eddy C, Wu L, Gamble GD, Mills GD, Garrett JE, Eaton TE, Rees MI. Functional variants of antioxidant genes in smokers with COPD and in those with normal lung function. Thorax 2006; 61:394–399.