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EDITORIAL

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Epidemiological, serological, and genetic evidence of influenza D virus infection in humans: Is it a justifiable cause for concern?

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Several new human viruses have emerged or re-emerged in recent years (e.g., SARS-CoV-2 [1–5], monkeypox virus [6], and Langya henipavirus LayV [7]) that have made significant global and/or local public health impacts. Yet, there are other viruses that appear to be spreading rapidly among livestock in recent years [8–10]. These newly emerged viruses can pose a significant economic burden on the agricultural sector and possibly to human health if they are left unchecked. The focus of this article will be on IDV, as mounting evidence has suggested that it has the potential to establish a recurring infection cycle in human populations, similar to that of influenza A (IAV) and influenza B (IBV) viruses, and might therefore require a seasonal immunization program, if it is proven necessary.

IDV was discovered in Oklahoma, USA in 2011 in an ailing pig [11]. Since then, it has been epidemiologically and serologically identified in cattle with respiratory disease [12]. Cattle appears to be the primary reservoir [13] for virus amplification and possible transmissions into other animal species [14,15], including swine, small ruminants, and camelids. Experimentally, IDV can infect and/or be transmitted to ferrets, mice, guinea pigs, cattle, and of course, cattle and pigs [11,13,16,17]. The ability of this virus to infect and/or transmit to multiple animal species raises a real concern about its zoonotic potential. Here, we summarize several lines of evidence that implicate IDV infection of humans and how to combat it.

A serological survey of sera collected from the general human population in the United States and Canada in 2011 showed a 1.3% seroprevalence of IDV in 4 out of 316 tested individuals [11] (Figure 1). A related and more comprehensive study in Italy found a relatively sharp increase in seroprevalence for IDV in its general human population from 5.1% in 2005 to 46% in 2014 [18]. Both studies linked serological evidence to environmental and occupational exposure to IDV. Since IDV has been identified in swine and cattle, it is important to assess the potential and degree of seroprevalence of workers in swine and cattle farms. It is remarkable that a very high level of seroprevalence (almost 100%, 34/35 IDV antibody-positive sera) was found among cattle workers in the United States [19]. However, a much lower degree of seroprevalence (4.9%, 4/82 IDV antibody-positive sera) was detected among swine veterinarians in Italy [20]. Regardless, these studies suggest that humans could be exposed to IDV under different settings (Figure 1), assuming that the potential for cross reactivity to other viruses (e.g., influenza C virus or ICV) could be ruled out, and highlight the significance of a potential for zoonotic transmission of IDV. However, it is important to note that, despite the serological evidence, there is no direct evidence that IDV can infect humans.

The closest and perhaps strongest evidence for a direct human infection by IDV came from a study, which showed that the IDV genomic materials could be detected in a nasal wash sample collected from a swine farm worker in Malaysia [21] (Figure 1). Additionally, IDV genomic materials have also been detected in aerosol samples collected from public settings, such as in an international airport [22], and in a hospital emergency room [23] in the United States. Collectively, these are corroborating evidence for zoonotic potential of IDV, which has the potential to pose a significant public health problem, like in the case of SARS-CoV-2 transmissions in public and/or congregate settings, including school, entertainment, hospice care and hospital settings, etc. [24-28]. Yet, it is important to note that, thus far, neither the complete viral sequence has been detected in those samples nor a replicationcompetent IDV has been isolated from them.

Experimentally, it has been shown that IDV can infect a variety of human cell types, including the human alveolar epithelial A549 cells, rectal tumor HRT-18 G cells [29,30], and other human primary well-differentiated airway epithelial cells [30], as well as

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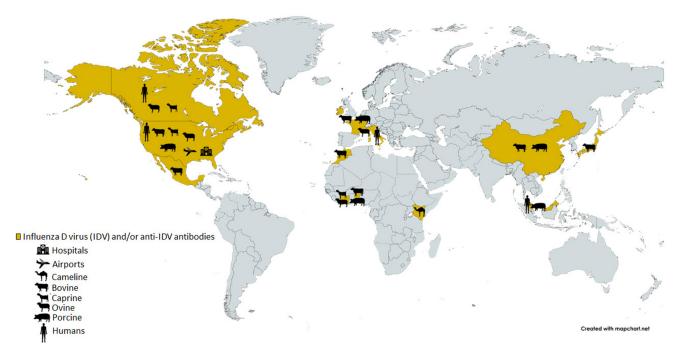


Figure 1. Locations where IDV or IDV antibodies have been detected, as well as the locations of environmental/occupational exposures are shown. Adapted from (14).

animal cells, including the Madin-Darby canine kidney MDCK cells, swine testicle ST cells, and the African green monkey kidney Marc-145 cells [11]. The ability of IDV to infect a variety of human and animal cells appear to be dependent on its ability to attach to and enter the cells via a common cellular receptor known as 9-O-acetyl-N-acetylneuraminic acid [29]. Additionally, the unique conformational structure of the IDV haemagglutinin esterase fusion (HEF) glycoprotein can partly explain its broad cellular tropism [29]. It is noteworthy that IDV has been shown to be able to experimentally infect and/or transmit to ferrets, mice, guinea pigs, and of course, pigs, all of which have been used as surrogate models of human influenza virus infection [11,13,16,17]. These findings highlight a real potential of IDV to infect and/or maintain not only in humans but also in other reservoirs, such as wild and domestic animals. As many of these animals have interacted with humans, it is important to perform routine serological surveys of individuals, such as swine and cattle workers, for evidence of IDV infections.

While IDV has not been shown to cause a significant form of illness in humans and therefore should not be of a significant public health threat, it has the potential to adapt and evolve (via spontaneous mutagenesis) in any of the susceptible hosts into a potentially more virulent form due to its error-prone RNA-dependent RNA polymerase that cannot faithfully copy its genome [31]. This has important public health implications in that some of the IDV natural variants might be able to establish its foothold in either the general or certain human population(s), such as the young or senior and/or immunocompromised individuals, and therefore could continue to evolve into different viral variant forms that might complicate the surveillance effort and/or might require seasonal immunization program, if an IDV vaccine is determined to be necessary and can be formulated as those of the currently available seasonal influenza quadrivalent vaccines [32–34] and therefore can be administered as a new IAV, IBV, and IDV combination vaccine formulation on an annual basis.

As the world is still grappling with the ongoing COVID-19 pandemic, it is important to keep viruses, such as IDV with zoonotic potentials, in check as some of these viruses have the potential to become human pathogens with deadly consequences.

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References

- Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet. 2020;395(10223):497–506. PMID: 31986264. DOI:10.1016/S0140-6736(20)30183-5
- [2] Zhou P, Yang XL, Wang XG, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature. 2020;579(7798):270–273. PMID: 32015507. DOI:10.1038/s41586-020-2012-7
- [3] Wu F, Zhao S, Yu B, et al. A new coronavirus associated with human respiratory disease in China. Nature. 2020;579(7798):265–269. PMID: 32015508. DOI:10.1038/s41586-020-2008-3
- [4] Zhu N, Zhang D, Wang W, et al. A novel coronavirus from patients with pneumonia in China, 2019. N Engl J Med. 2020;382(8):727–733. PMID: 31978945. DOI: 10.1056/NEJMoa2001017.
- [5] Lu R, Zhao X, Li J, et al. Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. Lancet. 2020;395(10224):565–574. PMID: 32007145. DOI:10. 1016/S0140-6736(20)30251-8
- [6] Centers for Disease Control and Prevention (CDC). Monkeypox. https://www.cdc.gov/poxvirus/monkey pox/response/2022/index.html
- Zhang X, Li H, Jiang FC, et al. A zoonotic henipavirus in febrile patients in China. New Engl J Med. 2022;387 (5):470-472. PMID: 35921459. DOI: 10.1056/ NEJMc2202705.
- [8] United States Department of Agriculture. (USDA). Animal Health Diseases. https://www.aphis.usda.gov/ aphis/ourfocus/animalhealth/animal-diseaseinformation/fmd/index
- [9] Huang C, Yu J, Hause BN, et al. Emergence of new phylogenetic lineage of Influenza D virus with broad antigenicity in California, United States. Emerg Microbes Infect. 2021;10(1):739–742. PMID: 33771071. DOI:10.1080/22221751.2021.1910078
- [10] Liu R, Sheng Z, Huang C, et al. Influenza D virus. Curr Opin Virolo. 2020;44:154–161. PMID: 32932215. DOI:10.1016/j.coviro.2020.08.004
- [11] Hause BM, Ducatez M, Collin EA, et al. Isolation of a Novel Swine Influenza Virus from Oklahoma in 2011 Which is Distantly Related to Human Influenza C Viruses. PLoS Pathog. 2013 Feb;9(2):e1003176. PMID: 23408893. DOI:10.1371/journal.ppat.1003176
- [12] Hause BM, Collin EA, Liu R, et al. Characterization of a novel influenza virus in cattle and swine: proposal for a new genus in the Orthomyxoviridae family. MBio. 2014 Mar 4;5(2):e00031. PMID: 24595369. DOI:10. 1128/mBio.00031-14.

- [13] Ferguson L, Olivier AK, Genova S, et al. Pathogenesis of influenza D virus in cattle. J Virol. 2016 Jun 15;90 (12):5636–5642. DOI:10.1128/JVI.03122-15.
- [14] Salem E, Cook EA, Lbacha HA, et al. Serologic evidence for influenza C and D virus among ruminants and camelids, Africa, 1991–2015. Emerg Infect Dis. 2017 Sep;23(9):1556. DOI:10.3201/eid2309.170342.
- [15] Zhai SL, Zhang H, Chen SN, et al. Influenza D virus in animal species in Guangdong Province, southern China. Emerg Infect Dis. 2017 Aug;23(8):1392. DOI:10.3201/eid2308.170059.
- [16] Sreenivasan C, Thomas M, Sheng Z, et al. Replication and transmission of the novel bovine influenza D virus in a guinea pig model. J Virol. 2015 Dec 1;89 (23):11990–12001. DOI:10.1128/JVI.01630-15.
- [17] Oliva J, Mettier J, Sedano L, et al. Murine Model for the Study of Influenza D Virus. J Virol. 2020 Jan 31;94
 (4): e01662. DOI:10.1128/JVI.01662-19 PMID: 31776281; PMCID: PMC6997775.
- [18] Trombetta CM, Marchi S, Manini I, et al. Influenza D virus: serological evidence in the Italian population from 2005 to 2017. Viruses. 2019 Dec 27;12(1):30. DOI:10.3390/v12010030
- [19] White SK, Ma W, McDaniel CJ, et al. Serologic evidence of exposure to influenza D virus among persons with occupational contact with cattle. J Clin Virol. 2016 Aug 1;81:31–33. DOI:10.1016/j.jcv.2016.05.017.
- [20] Trombetta CM, Montomoli E, Di Bartolo I, et al. Detection of antibodies against influenza D virus in swine veterinarians in Italy in 2004. J Med Virol. 2022 Jun;94(6):2855–2859. DOI:10.1002/jmv.27466. Epub 2021 Nov 27. PMID: 34811769; PMCID: PMC9299042.
- [21] Borkenhagen LK, Mallinson KA, Tsao RW, et al. Surveillance for respiratory and diarrheal pathogens at the human-pig interface in Sarawak, Malaysia. PLoS One. 2018 Jul 27;13(7):e0201295. DOI:10.1371/ journal.pone.0201295.
- [22] Bailey ES, Choi JY, Zemke J, et al. Molecular surveillance of respiratory viruses with bioaerosol sampling in an airport. Trop Dis Travel Med Vaccines. 2018 Dec;4 (1):1–5.
- [23] Choi JY, Zemke J, Philo SE, et al. Aerosol sampling in a hospital emergency room setting: a complementary surveillance method for the detection of respiratory viruses. Front Public Health. 2018 Jun 14;6:174. DOI:10.3389/fpubh.2018.00174.
- [24] Andrejko KL, Pry JM, Myers JF, et al. Effectiveness of face mask or respirator use in indoor public settings for prevention of SARS-CoV-2 infection—California, February–December 2021. Morbidity Mortality Weekly Rep. 2022 Feb 11;71(6):212. DOI:10.15585/ mmwr.mm7106e1.
- [25] Boehme C, Hannay E, Sampath R. SARS-CoV-2 testing for public health use: core principles and considerations for defined use settings. Lancet Glob Health. 2021 Mar 1;9(3):e247–9. DOI:10.1016/S2214-109X(21)00006-1
- [26] Macartney K, Quinn HE, Pillsbury AJ, et al. Transmission of SARS-CoV-2 in Australian educational settings: a prospective cohort study. Lancet Child Adolesc Health. 2020 Nov 1;4(11):807–816. DOI:10.1016/S2352-4642(20)30251-0.

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- [27] Zhou J, Otter JA, Price JR, et al. Investigating SARS-CoV-2 surface and air contamination in an acute healthcare setting during the peak of the COVID-19 pandemic in London. Clinl Infect Dis. 2020 Jul 8. DOI:10.1093/cid/ciaa905.
- [28] Birgand G, Peiffer-Smadja N, Fournier S, et al. Assessment of air contamination by SARS-CoV-2 in hospital settings. JAMA Network Open. 2020 Dec 1;3(12):e2033232. DOI:10.1001/jamanetworkopen.2020.33232.
- [29] Song H, Qi J, Khedri Z, et al. An open receptor-binding cavity of hemagglutinin-esterase-fusion glycoprotein from newly-identified influenza D virus: basis for its broad cell tropism. PLoS Pathog. 2016 Jan 27;12(1): e1005411. DOI:10.1371/journal.ppat.1005411
- [30] Holwerda M, Kelly J, Laloli L, et al. Determining the replication kinetics and cellular tropism of influenza D virus on primary well-differentiated human airway

epithelial cells. Viruses. 2019 Apr 24;11(4):377. DOI:10. 3390/v11040377

- [31] Kautz TF, Forrester NL. RNA virus fidelity mutants: a useful tool for evolutionary biology or a complex challenge? Viruses. 2018 Nov 1;10(11):600. DOI:10. 3390/v10110600
- [32] Belshe RB. The need for quadrivalent vaccine against seasonal influenza. Vaccine. 2010 Sep 7;28:D45-53. DOI:10.1016/j.vaccine.2010.08.028.
- [33] Ambrose CS, Levin MJ. The rationale for quadrivalent influenza vaccines. Hum Vaccin Immunother. 2012 Jan 1;8(1):81–88. DOI:10.4161/hv.8.1.17623
- [34] Centers for Disease Control and Prevention (CDC). Prevention and control of seasonal influenza with vaccines. Recommendations of the Advisory Committee on Immunization Practices–United States, 2013-2014. MMWR Recomm Rep. 2013 Sep 20;62(RR–07):1–43.